

Stephen G. Williams · John P. Stein

Molecular pathways in bladder cancer

Received: 16 May 2001 / Accepted: 30 June 2003 / Published online: 13 November 2004
© Springer-Verlag 2004

Abstract The aim of this review is to provide a contemporary outline of our current understanding of the molecular and genetic events associated with tumorigenesis and the progression of bladder cancer. A comprehensive review of the literature was performed on the molecular alterations associated with transitional cell carcinoma (TCC) of the bladder. Intense research efforts are being made to better identify and characterize various bladder cancers and their true biologic potential. The need to predict which superficial tumors will recur or progress, and which invasive tumors will metastasize has led to a much better understanding of the molecular pathways associated with bladder cancer. The molecular changes that occur in TCC of the bladder are numerous and can be categorized into: (1) chromosomal alterations leading to carcinogenesis, (2) loss of cell cycle regulation accounting for cellular proliferation, and (3) metastasis, guided by events such as angiogenesis. It is becoming apparent that the accumulation of genetic and molecular changes ultimately determines a tumors phenotype and subsequent clinical behavior. At the present time, conventional histopathologic evaluation of bladder cancer (tumor grade and stage) is inadequate to accurately predict the behavior of most bladder tumors. While new laboratory techniques have allowed us to better understand how bladder cancer develops and ultimately progresses, few of these techniques are currently available for use in the clinical setting. The ultimate goal is to develop reliable prognostic markers which will accurately predict

not only the expected clinical course of an individual bladder tumor but also the response of that tumor to currently available therapies. More importantly, this information may be employed in the future to dictate altogether new treatments for the prevention and/or stabilization of the early molecular events that lead to the development of bladder cancer.

Keywords Molecular pathway · Genetic events · Bladder cancer · Tumorigenesis · Transitional cell carcinoma

Transitional cell carcinoma (TCC) of the bladder is the second most common malignancy of the genitourinary tract, and the second most common cause of death of all genitourinary tumors. In 2003, there were 42,200 new cases of the disease diagnosed, with 8,600 patient deaths [1]. Approximately 80% of patients with primary bladder cancer present with low grade tumors confined to the superficial mucosa. The risk of recurrence in these patients with superficial bladder tumors can be as high as 70%, with the majority of cancers amenable to initial transurethral resection and the selected administration of intravesical immuno- or chemotherapy [2]. Unfortunately, as many as 30% of these recurrent tumors may demonstrate tumor progression to a higher grade and/or stage of disease. Furthermore, 15–30% of all patients with bladder cancer initially present with muscle invasive tumors, 50% of these patients who are treated locally for their invasive tumors will relapse with metastatic disease within 2 years of treatment [3]. These data underscore the heterogeneous nature and malignant potential of TCC of the bladder.

The optimal management of invasive bladder cancer requires the detection and accurate assessment of the tumor's biologic potential. Currently, histologic evaluation, including determination of tumor grade and stage, is the primary prognostic variable which dictates treatment strategies for patients with bladder cancer. Although these two conventional histopathologic variables

S. G. Williams (✉)
Department of Urology, Kaiser Permanente Medical
Center - Los Angeles, 4900 Sunset Blvd,
Los Angeles, CA 90027, USA
E-mail: stephen.g.williams@kp.org
Tel.: +1-323-783-5500
Fax: +1-323-783-7272

J. P. Stein
Department of Urology,
Kenneth Norris Jr. Comprehensive Cancer Center,
1441 Eastlake Ave., Suite 7416, Los Angeles, CA 90089,
University of Southern California, USA

provide a certain degree of stratification of a tumor's biologic potential, there remains a significant degree of tumor heterogeneity even within various prognostic subgroups. This makes the accurate and reliable prediction of the tumor's aggressiveness difficult. The ability to precisely predict an individual tumor's true biologic potential would in turn facilitate treatment selection decisions for patients who may benefit from adjuvant therapy, and identify patients who may require less aggressive treatment strategies. Intense research efforts are ongoing to identify and better characterize bladder cancer and its varying biologic potential.

TCC of the bladder has generally been viewed as two different disease processes. Superficial bladder tumors are thought to be more of a locally proliferative, recurrent process, but can become invasive and even metastatic. The use of molecular markers may guide decision making processes in the treatment of superficial bladder cancer [4]. Superficial bladder tumors that maintain a malignant phenotype may be better treated with early, aggressive intravesical therapy, or even cystectomy. On the other hand, muscle invasive bladder cancer is notorious for its potential clinical virulence and is ideally treated aggressively [2]. Despite this aggressive form of therapy, there remains a significant incidence of recurrence and disease progression in patients, who may ultimately benefit from some adjuvant form of therapy.

The need to predict which superficial bladder tumors will recur or progress, and which invasive tumors will metastasize, has led to the ongoing attempt to understand bladder carcinogenesis and metastasis. With the advent of new molecular techniques, the field of medical molecular biology has exploded in recent years, resulting in detailed analyses of human cells and tissues at the DNA, RNA and protein levels. The molecular and genetic changes in TCC of the bladder can be schematically classified into three separate, but intertwined events: (1) chromosomal alterations—representing the initial event in carcinogenesis, (2) tumor proliferation—due to loss of cell cycle regulation, and (3) metastasis—in which the initial tumor breaks from its original, confined environment aided, in part, by processes such as angiogenesis and the loss of cell adhesion. We believe it is the accumulation of these successive genetic alterations, rather than a single genetic event in time, that determines a tumor's phenotype and, subsequently, the patients clinical outcome. In this review, we will summarize the recent literature on the molecular and genetic changes in bladder cancer and comment on potentially improved diagnostic abilities and treatment regimens that are becoming available as a result of our improved understanding of these molecular pathways.

Initiation of carcinogenesis—oncogenes and chromosomal alterations

Bladder cancer is an excellent model for the study of molecular changes at the DNA level, due to its distinctly

different subtypes—superficial and muscle-invasive—and their different propensities to progress. Such DNA alterations in bladder cancer have been studied in a variety of ways, ranging from cytogenetics to DNA ploidy to loss of heterozygosity (LOH) [5]. DNA alterations can result from any number of genetic insults such as mutations (point and insertional/deletional), translocation, and loss of alleles. Each insult may effect the translated protein product. The large fund of molecular knowledge on carcinogenesis, developed in recent years, has provided some evidence of the different genetic pathways for bladder cancer.

Earlier work in the field of molecular oncology focused on oncogenes. Oncogenes are normal cellular genes that contribute to the malignant phenotype of a tumor by overexpressing the normal gene product, or, in some cases, by expressing a protein product with an altered function. Overexpression of the normal gene product is usually achieved by gene amplification or chromosomal translocation of the gene to an area downstream of a powerful promoter. However, the expression of a mutated protein product can also lead to activation of the malignant phenotype. Oncogenes believed to be important in human malignancies include: c-H-ras, c-myc, mdm2, and c-erbB2.

c-H-ras

The c-H-ras gene is an active oncogene thought to be involved in the development and progression of human bladder cancer. Mutational studies of the ras gene family have demonstrated that alterations in codon 12 and 61 of the H-ras gene occur in up to 20% of bladder cancers [6, 7, 8]. One study employing polymerase chain reaction amplification followed by oligonucleotide specific hybridization reported that 36% of bladder tumors had the same mutation at codon 12 of the H-ras gene [9]. In general, the activation of H-ras occurs by a single point mutation (G to A) in condon 12, although other mutations have been described [6]. Clinically, Fontana and colleagues demonstrated a statistically significant relationship between the overexpression of the c-ras oncogene and early recurrence in patients with superficial bladder cancer [10]. These data suggest a potential prognostic role for the c-ras oncogene in patients with superficial bladder cancer, but currently these techniques apply only in a research setting.

c-myc

The myc gene family is an important regulator of cellular proliferation and encodes for nuclear phosphoproteins containing DNA-binding activity [11]. The c-myc oncogene is overexpressed in several human tumors including bladder cancer [12, 13]. Deregulation of the myc gene family occurs with chromosomal translocation and gene amplification [14], and studies have demonstrated that

myc overexpression promotes cellular proliferation [10]. Although the genetic mechanism causing overexpression of the c-myc gene in bladder cancer is unknown, overexpression of c-myc has been shown to be associated with high grade bladder cancer. Kotake and associates demonstrated that expression of the c-myc gene product correlates with the nuclear grade of bladder cancer [13]. In a conflicting study, Lipponen found no independent prognostic value for myc proteins with respect to the prognosis of patients with TCC of the bladder [15]. Currently, the prognostic significance of c-myc gene expression is unknown and further evaluation will be required to determine its prognostic role.

c-erbB-2

The proto-oncogene c-erbB-2 (also known as HER-2/neu) has been extensively studied and implicated in a number of tumors including breast, prostate and bladder cancer [16]. The c-erbB-2 oncogene encodes a transmembrane glycoprotein, similar to the epidermal growth factor receptor, having tyrosine kinase activity [17] and the ability to stimulate cellular growth [18]. Initial studies of c-erbB-2 were performed in breast carcinoma, and demonstrated a significant relationship between gene expression, tumor progression and overall survival [19]. Subsequently, several studies have reported that c-erbB-2 expression in patients with bladder cancer is associated with higher stage tumors [20, 21, 22], increased tumor progression [16], increased incidence of metastasis [22], and decreased overall survival [20]. Although these studies suggest a prognostic value of c-erbB-2 expression in human bladder cancer, other studies have reported conflicting results, concluding that the evaluation of c-erbB-2 provides no additional prognostic value over previously established predictors (grade and stage) for TCC of the bladder [23, 24]. In view of these discrepant results, further evaluation will be required to accurately determine the prognostic value of c-erbB-2 in bladder cancer.

Tumor suppressor genes

More recent work in the field of chromosomal alterations has focused on identifying specific loci on chromosomes which may contain altered genes. Many of these genes have been identified as tumor suppressor genes (TSG) that, when inactivated, result in the initiation and/or progression of the malignant phenotype. Recently, with the advent of such techniques as LOH analysis and comparative genomic hybridization (CGH), a significant increase in genome scanning has occurred, with the identification of many new chromosomal alterations in TCC. LOH analysis uses known polymorphic markers to identify large deletions and/or alterations of both alleles of a chromosome. The most common scenario would be one large deletion of an

entire chromosome due to natural genetic recombination, followed by a small alteration in the retained allele, involving an insertional, deletional, or point mutation, usually at a specific locus containing a tumor suppressor gene. CGH utilizes genomic DNA from tumor and normal cells that is differentially labeled by fluorescence. The two DNA extracts are then hybridized on to a platform of normal metaphase spreads (of all human chromosomes). A loss or amplification of a particular DNA sequence on the tumor DNA is determined by comparing label intensity to the normal hybridized DNA.

Using these molecular techniques, an extensive search in recent years has led to finding several key TSGs on different chromosomes. Deletions on the short arm of chromosomes 3 (3p) [25] and 8 (8p) have been found with high grade, muscle-invasive bladder cancer. In fact, the 8p deletion has been noted in >50% of muscle-invasive TCC [26, 27], (a rate similar to that of p53 mutations), while no deletions were noted in superficial TCC. LOH on the short arm of chromosome 8 (8p) has also been associated with high grades and stages of TCC, but the relevant gene(s) have not yet been identified. However, the most notable chromosomal deletions in bladder cancer have been found on chromosomes 9, 13, 17. This has led to the identification of the retinoblastoma (Rb) TSG on chromosome 13, the p53 TSG on chromosome 17, and promising new TSG on chromosome 9 at the p21 locus.

Deletions on chromosome 9 are the most common chromosomal abnormalities in TCC, and are found in >50% of all grades and stages of TCC. However, it is becoming clear that most muscle invasive bladder tumors have other chromosomal alterations as well. On the other hand, most Ta and T1 tumors show few genetic alterations other than on chromosome 9 [28, 29, 30]. This has led to the hypothesis that inactivation of genes on chromosome 9 may represent an early event in the development of bladder cancer.

The majority of deletions on chromosome 9 have been found on the short arm (9p). Specifically, a complex genomic region at 9p21 (INK4A/ARF and INK4B) exists, encoding three distinct proteins—p16, p14ARF, and p15—all of which act as negative cell cycle regulators and are considered potential TSGs [31]. Extensive screening of the retained allele in bladder tumors with 9p LOH have not revealed frequent mutations. Instead, deletion of both alleles (homozygous deletion) appears to be the common mechanism of inactivating the entire locus in TCC [32, 33, 34]. Such a deletion will commonly remove all three 9p21 genes. While the clinical implications of such homozygous deletions have not been evaluated, in vitro transfection studies using bladder cancer cell lines have shown growth arrest following the introduction of p16 into cell lines with deleted INK4A [35]. Identification of the p14ARF, p15, and p16 proteins, for use as markers of malignant potential, are currently under investigation, and will be discussed further in the section on cell cycle regulation.

Recent studies involving large LOH analyses have also identified possible loci for TSG on 9q, at 9913–31, 9932–33 and 9934 [75, 76, 80]. Candidate genes have been identified at 9932–33 (deleted in bladder cancer candidate region 1-DBCCR1) and 9934, the same region as the tuberous sclerosis gene. The DBCCR1 gene shows no significant homology to other known genes, and is expressed in the urothelium of adult bladder and ureter. However, in one study about half of the bladder tumor cell lines studied showed no expression of DBCCR1, perhaps indicating gene silencing secondary to DNA methylation in the promoter region [36]. Most investigators have found no relationship between 99 loss and tumor invasiveness, possibly supporting its role in the initiation of bladder cancer, rather than in progression [37].

Two other very important chromosomal alterations that affect known TSGs involve 17p13, the site of the p53 gene, and 13914, the site of the Rb gene. The importance of p53 in bladder TCC was suggested by the high frequency of LOH of chromosome 17p in high grade TCC [38]. Others have confirmed that p53 follows classic tumor suppressor theory, with LOH at one allele (17p) and a mutation of the remaining TP53 allele. This pattern has subsequently been identified in a large number of muscle-invasive bladder tumors, with a lower frequency in superficial tumors [39, 40]. These genetic defects have been demonstrated to correspond with protein expression of the mutated p53 gene product [41, 42]. However, p53 evaluation remains a good example of the difficulty of making the transition from genetic studies to translational research techniques such as immunohistochemistry (IHC). Nearly 25% of tumors that are p53-altered (positive) by IHC show no detectable mutations by standard gene sequencing analyses, while about 10% of wild-type tumors (p53 negative) harbor readily identifiable mutations [42, 43]. Still, p53 remains the best characterized TSG to date and is clearly implicated in disease progression in several solid tumors, including bladder cancer.

The Rb gene was the first TSG isolated. It codes for a nuclear protein mapping to 13914 [44]. Like p53, Rb is thought to play an important role in bladder cancer progression. Rb gene mutations are noted in 25–30% of bladder tumors [45, 46], and LOH at the Rb locus (13q) is strongly associated with the absence of Rb protein expression by IHC [47]. However, unlike p53, missense mutations are rare in RB and no mutational hotspots have been identified. In fact, mutations have been found distributed throughout 24 of the 27 exons in hereditary retinoblastoma. Therefore, detailed analyses of specific mutations within the Rb gene are unlikely to be used in clinical decision making as the large size of the gene does not lend itself to standard DNA sequencing or single strand conformational polymorphism analysis [48].

In a recent study, 12 of 19 bladder tumor cell lines had identifiable mutations in exons 5–11, considered the central domain, of TP53, with a concomitant loss of Rb protein expression. The other seven bladder cancer cell

lines studied showed wild-type p53 (normal) or mutations only in exons 1–4, but all seven cell lines had concomitant alterations at the 9p21 (INK4A/ARF and INK4B) gene locus [49]. This study provided the first evidence for possible differences in the penetrance of p53 mutations in bladder cancer, while adding evidence to the concept of multiple genetic pathways of bladder carcinogenesis, first proposed by Spruck et al. [50].

We have known for years that superficial and muscle-invasive TCC of the bladder are quite different histologically and that they behave altogether differently in the clinical realm. On the basis of the findings described above, of frequent and consistent genetic alterations in the two primary forms of bladder cancer, we are now able to come up with a fairly detailed model of these “multiple genetic pathways of bladder carcinogenesis” (Fig. 1). The key studies to date indicate that chromosomal instability leading to alterations in cell cycle regulation are integral events in determining the biologic behavior of bladder tumors. Although our understanding of the actual molecular events leading to the initiation of bladder cancer is growing rapidly, none of the techniques currently available to identify specific allelic instability or LOH in individual bladder tumors are yet applicable for clinical use. The advent of high-throughout molecular technology, however, may allow for clinical testing of specific chromosomal alterations in individual patients in the near future.

The cell cycle regulatory pathway

Normal cellular proliferation occurs by an orderly progression through the cell cycle which is regulated by cell cycle associated protein complexes composed of cyclins and cyclin-dependent kinases [51]. Several TSGs acting

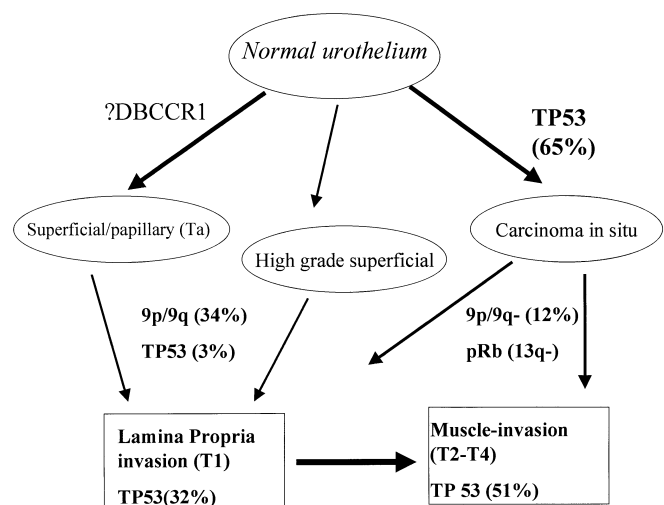


Fig. 1 Molecular model of bladder cancer and its progression showing key molecular alterations that have been described. Values in parentheses for chromosome 9 and TP53 mutation frequencies are from [50]. Adapted from [31]

at the G0/G1 checkpoint of the cell cycle are now recognized, and their protein products—p53, pRb, p16, and p14—are vital for preventing cell cycle progression in bladder tumors. Inactivation of one or more TSGs and loss of cell cycle control appear to be early steps in the development of carcinogenesis and ultimately cancer progression. Inactivation of a gene can occur by mutation, deletion, or methylation, and in most cases requires alteration of both copies of the gene: p53 is an exception as the alteration of only one copy of the gene is sufficient to alter function. The inactivation of both copies of a gene can occur by one of two pathways: (1) primary inherited alteration of one copy, followed by a second “hit” occurring somatically (due to environmental mutagen exposure or dysfunction of DNA replication/repair), or (2) an entirely somatic event(s) in which two independent “hits” occur in both copies of a gene [48].

While most of the initial studies on cell cycle regulation in bladder cancer focused on the role of individual TSG and their protein products, more recent investigative efforts have identified multiple pathways within the cell cycle. These pathways involve the interaction of multiple TSG, and it is this interaction that is most likely responsible for bladder cancer progression.

Retinoblastoma TSG

The Rb gene is located on chromosome 13p14 and encodes for a 110 kDa nuclear phosphoprotein [52]. Although initially discovered to be mutated in patients with inherited retinoblastoma, altered Rb gene expression has been reported in various human tumors including TCC of the bladder [45, 46, 53]. In its physiologic active hypophosphorylated form, pRb acts by inhibiting cell cycle progression at the G1-S checkpoint. However, pRb interacts with multiple cell cycle regulatory proteins, including: (1) cyclins, which catalyze the inactivation of pRb via phosphorylation, (2) cdk inhibitors, including p21, p16, and p27, which activate cdk/cyclin complexes, thus inhibiting pRb phosphorylation, and (3) the E2F family of transcription factors, which are responsible for transactivating genes necessary for entry into the S (synthesis) phase of the cell cycle [54]. Any alteration in those interactions can lead to uncontrolled cell growth.

Inactivation of the Rb gene is thought to be an important step in bladder cancer progression. With a combination of immunohistochemical techniques and molecular analysis, several groups have demonstrated that the proportion of tumors demonstrating Rb alterations increases with higher grade and stage bladder cancers [47, 55]. The results of these studies suggest that the loss of pRb expression may be an important prognostic factor in TCC of the bladder. Cordon-Cardo and associates reported that patients with muscle invasive bladder tumors who had lost Rb immunoreexpression had a significantly shorter 5-year survival than those patients which normal Rb protein expression [46]. Similarly,

Logothetis and associates studied 43 patients with invasive bladder cancer and demonstrated that Rb alterations were more common in advanced tumors, and that those patients who had lost pRb expression had a shorter overall survival compared to those who had maintained Rb expression [45]. Based on the aforementioned data, it appears that pRb expression is an important prognostic factor in patients with invasive bladder cancer.

p53 TSG

Mutations in the p53 gene are the most common genetic defect in human tumors [56]. The p53 gene is located on chromosome 17p13 and encodes for a 53-kDa protein. The p53 gene is known to play a vital role in the regulation of the cell cycle [57]. When DNA damage occurs, the level of p53 protein increases causing cell cycle arrest. This allows for the repair of DNA and prevents propagation of the DNA defect. Mutations in the p53 gene result in the production of an abnormal and usually dysfunctional protein product with a prolonged half-life compared to the wild-type protein. Consequently, this abnormal protein accumulates in the cell nucleus and can be detected by immunohistochemical staining. Several studies have demonstrated that the nuclear accumulation of p53 protein, as determined by immunohistochemical staining, correlates with gene mutations detected by DNA sequence analysis [42, 58, 59].

p53 alteration, as determined by immunohistochemical techniques, is an important prognostic indicator for bladder cancer progression [41, 60, 61]. Increased p53 immunoreactivity has been found in higher grade and stage bladder cancers and is associated with disease progression and decreased overall and disease-specific survival. Our group evaluated p53 nuclear reactivity in 243 patients with invasive bladder cancer who were uniformly treated by radical cystectomy [60]. Patients with an increased p53 expression (altered p53) were found to have a significantly increased risk of disease recurrence and a significantly decreased overall survival when compared to those patients without altered p53. This association was strongest in patients with organ confined bladder tumors (P1, P2, P3a). Furthermore, nuclear accumulation of p53 was found to be the only independent predictor of disease progression in a multivariate analysis of p53 status, histologic grade and pathologic stage.

Evidence is now accumulating that the mutation status of p53 varies greatly, and that not all p53 mutations affect the cell cycle in the same manner. Most studies of p53 mutations have identified alterations in exons 5–8, the central core domain of p53 [3]. However, the frequency of mutations outside this region (exons 2–4 and 9–11), although lower, is highest in bladder cancer compared to other cancers [62]. Markl and Jones have shown, in bladder cancer cell lines, that mutations in

exon 4 of p53 were always associated with loss of p14/p16, while mutations in exons 5–11 were always paired with loss pRB [49].

Combination of Rb and p53 TSGs

Two independent studies have evaluated the prognostic significance of combining the Rb and p53 status of bladder cancers as determined by immunohistochemical techniques [63, 64]. Preliminary data from these studies support the concept that bladder tumors with alterations in both p53 and Rb have a poorer prognosis and decreased overall survival when compared to tumors with wild-type p53 and wild-type Rb. Tumors with an alteration of only one of these genes (as determined by immunohistochemistry) behave in an intermediate fashion. These data suggest that the status of both p53 and Rb are important, and that these two proteins act in an independent yet synergistic manner in patients with bladder cancer.

p21 TSG

Although p53 nuclear accumulation, as detected by immunohistochemical methods, is a significant predictor of bladder cancer progression, not all p53-altered bladder tumors recur or progress [61, 60]. One of the primary functions of p53 is as a cell cycle regulatory protein [59]. p53 mediates its effects on the cell cycle, in part, through the regulation of p21^{WAF1/CIP1} expression [51]. Alterations in p53 can result in loss of p21 expression, which leads to unregulated cell growth. This is thought to be one of the mechanisms through which p53 alterations may influence tumor progression. However, it has recently been demonstrated that p21 expression may also be mediated through p53-independent pathways [65, 66]. This important finding suggests that despite the presence of a p53 alteration, p21 expression (and therefore cell cycle control) can be maintained.

We have evaluated bladder tumors from 101 patients who underwent radical cystectomy for invasive bladder cancer for p21 expression using immunohistochemical techniques [66]. All patients had been previously determined to have p53 altered tumors [60]. We found that immunohistochemical detection of p21 protein in the nuclei of bladder cancers which show p53 alterations (p53-altered) provides important additional prognostic information for patients with bladder cancer. Patients with p53-altered TCC of the bladder that were p21-negative demonstrated a significantly increased probability of recurrence and a significantly decreased probability of overall survival when compared to patients with p53-altered tumors that maintained expression of p21 (p21-positive). The association between p21 status and prognosis in p53-altered bladder tumors was independent of tumor grade, pathological stage and lymph node status. Loss of p21 expression was strongly

associated with an increased probability of recurrence and decreased probability of survival in patients with lymph node negative organ confined disease and lymph node negative extravesical disease. These findings suggest that p21 expression through p53 independent pathways exist, and that cell cycle control may be maintained through these pathways. Those patients with p53 altered tumors that lose p21 expression appear to have a poor prognosis and may best be managed with aggressive forms of therapy.

Deciphering cell cycle regulation in TCC

The interaction of p53 and p21 in cell cycle regulation, and the data looking at the cooperative effects of p53 and Rb, provide good examples of the increasing evidence that mutation in a single TSG is unlikely to be the only factor resulting in carcinogenesis. We now understand that there are several pathways within the cell cycle, each playing a role in cell cycle regulation. The alteration of one or more of these pathways is likely responsible for bladder cancer progression.

As discussed previously, the 9p21 locus is a complex region of chromosome 9, where many deletions have been identified in bladder tumors and bladder cancer cell lines. This locus has proven crucial in the regulation of the cell cycle because of the unusual situation in which two functionally different genes—p16 and p14—are both transcribed from the same locus, but via alternative first exons and reader frames.

p16 (also known as INK4A, MTS1, CDKN2A) is a well-characterized CDK inhibitor [67] which functions upstream of pRb to block cyclin-D directed phosphorylation of Rb, thus inducing G1 arrest. p16 mutations and homozygous deletions are common in bladder cancer cell lines and in squamous cell carcinoma and bilharzias associated bladder cancers [54]. Furthermore, p16 is thought to be susceptible to transcriptional silencing by promoter methylation [68]. Inactivation of p16 by any of the mechanisms will lead to uninhibited phosphorylation of pRb and subsequent cell proliferation.

The other gene product at 9p21 is p14, also known as ARF (alternative reading frame), and as p19 in the mouse. p14 acts upstream of p53 to stimulate p21 expression, and may also play an important role in the feedback loop that regulates the cellular level of p53 by interacting with the cellular proto-oncogene product MDM2 [69]. However, p14 is a very different TSG than p16 in that it is expressed ubiquitously, whereas p16 expression is more restricted; p14 does not bind to CDKs or function as a CDK inhibitor [70], and it can cause cell cycle arrest at any point in the cycle through its effect on p21 [48].

While the majority of research on the 9p21 locus has been performed in vitro using cell lines and animal models, Orlov et al. recently examined deletions of the INK4A gene in 121 patients with superficial (Ta or T1)

bladder tumors. They found that homozygous deletions of the INK4A gene resulted in a lower recurrence-free survival. Furthermore, deletions that affected both p16 and p14 (thus deregulating both the p53 and pRb pathways), correlated with larger and higher grade tumors [71].

As previously mentioned, p14 regulation of the cell cycle can occur through physical interaction with MDM2 [72]. In normal cells, MDM2 regulates p53 function by marking p53 for degradation via ubiquitin conjugation and inactivating p53 by binding to its transactivation domain [73]. p14 binding of MDM2 appears to counteract the effects of MDM2 by protecting p53 from degradation [73]. Nevertheless, the role of MDM2 in regulating p53 protein levels in TCC of the bladder remains unclear. The frequency of MDM2 gene amplification across all malignancies is thought to be uncommon (~7%), with the highest frequency in soft tissue tumors (20%). MDM2 amplifications and p53 mutations usually do not occur within the same tumor sample, indicating that carcinogenesis can result from MDM2 amplification alone. However, MDM2 gene amplification is infrequent in bladder cancer, with one study showing only one of 87 cases of MDM2 amplification, despite elevated MDM2 protein levels in 26 of the 87 cases [74].

Thus, the role of TSGs in cell cycle regulation is a complex one, with the 9p21 gene locus lying at the center of the two major tumor suppressor pathways identified in bladder cancer—p53 and Rb. The two pathways and the various TSG interactions described above are summarized in Fig. 2.

Angiogenesis

Angiogenesis, the formation of new blood vessels from the surrounding established vasculature, is a tightly regulated, essential physiologic process that occurs during normal development, reproduction and repair. Uncontrolled angiogenesis can lead to a variety of pathologic states and participates in the maintenance of neoplastic conditions. In its simplest form, angiogenesis can be described in three steps: (1) initiation and activation of the endothelial cells, (2) migration and invasion of the activated endothelial cells following proteolytic degradation of the surrounding extracellular matrix, and (3) maturation of the endothelial cells to coalesce and form water tight tubules that establish new blood flow [75]. Understanding this complex process in an attempt to inhibit tumor angiogenesis has been the focus of expanding interest and investigation in the field of oncology because of its potential therapeutic benefits.

Stimulation of angiogenesis (neovascularization) is a critical adaptation characteristic of all solid tumors. Without angiogenesis tumor growth is inhibited at a diameter of 2–3 mm, the natural limit for diffusion of essential nutrients and oxygen [76]. Under most homeostatic conditions, new blood vessel formation is

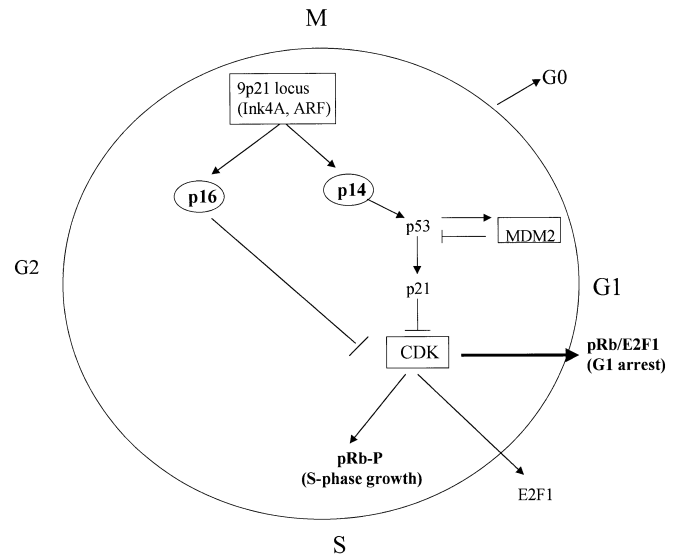


Fig. 2 The INK4A/ARF locus and cell-cycle regulation. The 9p21 locus yields two unique proteins, p14 and p16, which act along separate pathways of cell cycle control. p14 modulates expression of p53 by directly stimulating expression or indirectly downregulating expression via MDM2. p53 then upregulates expression of p21, inhibiting the cyclin-dependent kinase (CDK)-cyclin complex from phosphorylating pRb and leading to G1-phase arrest. The p16 pathway leads to direct inhibition of the CDK-cyclin complex, yielding cell cycle arrest through inhibition of pRb phosphorylation

infrequent and is controlled by an abundance of inhibitory signals directed at the endothelium, setting the balance in favor of vascular quiescence. It is thought that the angiogenic phenotype for any given tumor is determined by the overall balance between stimulatory and inhibitory inputs to the endothelial cells. During disease states such as carcinogenesis, the angiogenic balance within the tumor's microenvironment shifts in favor of endothelial cell activation. Folkman has termed this the "angiogenic switch", whereby new vessel growth is stimulated, thus providing the necessary nutrients for continued tumor growth and eventual metastasis [77]. Current research has identified several mechanisms by which this angiogenic switch can occur, including overexpression of inducers and/or loss of endogenous inhibitor production [78]. These factors may be produced by the tumor cells themselves or released from the surrounding extracellular matrix and tumor-associated stromal cells, or they may be products of inflammatory cells that infiltrate the tumor [79, 80]. As both tumor growth and invasion depend, in part, on this angiogenic response, the ability to quantitate the degree of angiogenesis within or around a given tumor may provide prognostic information. This has been accomplished by determining the so-called "microvessel density" within and around a given tumor using antibodies to Factor VIII and CD 34, which recognize immature or new vascular endothelial cells.

Microvessel density has been demonstrated to be a useful prognostic indicator in a variety of malignancies

including melanoma [81], breast cancer [82], and prostate cancer [83]. In general, increased microvessel density counts have been associated with tumor progression and decreased survival [83, 84]. The relationship between microvessel density count and tumor progression has also been examined in patients with bladder cancer [85, 86]. Dickinson et al. evaluated a series of 45 patients which invasive bladder tumors with a median follow-up of 37 months, and found microvessel density count to be an independent prognostic indicator of disease progression. Patients which an elevated microvessel density count demonstrated a 2.5-fold greater risk of dying [87].

Our group recently evaluated the relationship between tumor angiogenesis and tumor progression in 164 patients with invasive bladder cancer [85]. In this study, microvessel density was significantly associated with both disease recurrence and overall survival in these patients following radical cystectomy. Patients with elevated microvessel density counts demonstrated a significantly increased risk of disease recurrence and a worse overall survival when compared to patients with low microvessel density counts. Furthermore, microvessel density count was found to be an independent prognostic indicator of both disease progression and overall survival when evaluated in the presence of histologic grade, pathologic stage and regional lymph node involvement.

Angiogenic inducers

The difficulty in evaluating the angiogenic potential of any given tumor results from the abundance of pro-angiogenic factors that are produced by tumor cells or released by the surrounding extracellular matrix (ECM) [88] (Table 1). Prevailing evidence for the balance hypothesis proposed by Folkman suggests that an angiogenesis suppressor gene(s) encodes or controls the expression of one (or several) angiogenesis inhibitors that maintain a quiescent vasculature in cells. The theory maintains that the angiogenic inhibitor is downregulated during tumorigenesis, resulting in a balance in favor of the angiogenic inducers, with subsequent endothelial cell proliferation and migration (thus the term “angiogenic switch”) [77].

The angiogenic properties of urine from patients with TCC were first noted by Chodak and associates who documented a stimulatory effect on the migration of

endothelial cells exposed to the urine [89]. Basic fibroblast growth factor (bFGF), a potent pro-angiogenic factor, is excreted at higher levels in the urine of bladder cancer patients compared to patients without evidence of disease [90, 91]. Urinary bFGF has also been correlated with the pathologic stage of the primary tumor in patients with muscle-invasive TCC [92].

Recently, elevated levels of another important angiogenic inducer—vascular endothelial growth factor (VEGF)—have been found in the urine of bladder cancer patients. Crew and colleagues evaluated 98 patients with bladder cancer and found VEGF levels to be highest in the urine of patients with bladder cancer compared to normal controls and patients with other unrelated malignant conditions. In addition, they found that VEGF levels correlated with tumor recurrence in patients with Ta and T1 disease [93]. We recently found elevated levels of VEGF in the urine of bladder cancer patients with high grade and/or muscle-invasive TCC, as compared to patients with prostate cancer and patients without evidence of malignancy. In 92 patients undergoing radical cystectomy for muscle-invasive or high grade superficial TCC of the bladder, higher VEGF levels in urine obtained preoperatively were associated with which significantly decreased 3 year survival [94].

While numerous pro-angiogenic factors have been identified in bladder cancer cell lines and tissue, Campbell and colleagues have shown that VEGF and bFGF appear to be two primary inducers of angiogenesis in bladder cancer cell lines [88]. Neutralizing antibodies to VEGF, and to a lesser extent bFGF, significantly reduced the angiogenic activity of bladder cancer cell lines, whereas neutralizing antibodies to aFGF, scatter factor, TGF alpha and beta, and thymidine phosphorylase did not.

O'Brien and colleagues found that tissue levels of VEGF taken from human bladder cancer tumors correlated with the stage progression in superficial TCC. They found a fourfold increase in VEGF mRNA levels in Ta tumors compared to normal urothelium, and a tenfold increase in T1 tumors. Interestingly, T1 tumors also had an increased expression of VEGF mRNA when compared to invasive (T2–T4) tumors [95]. Other researchers have found a similar expression of VEGF in normal urothelial tissue and bladder cancer. Campbell et al. found relatively constant levels of VEGF immunostaining across normal urothelium, superficial and muscle-invasive bladder cancer [88]. Sato et al. found that the VEGF transcript was present in both normal urothelium and bladder cancer tissue. However, muscle-invasive tumors expressed significantly higher levels of VEGF by Northern blot analysis [96].

In most cases, angiogenic factors appear to be produced directly by the bladder cancer cells. Interestingly, the immunostaining pattern for bFGF appears to be unique in that bFGF localizes primarily to the basement membrane rather than tumor cells. O'Brien and colleagues hypothesized that tumor-induced degradation of the basement membrane could release bFGF, accounting

Table 1 Proangiogenic factors identified in bladder cancer

Proangiogenic factors	Acidic fibroblast growth factor [119, 120]
	Basic fibroblast growth factor [90, 92]
	Vascular endothelial growth factor [88, 95, 121]
	Thymidine phosphorylase [95, 122]
	Scatter factor [123]
	TGF beta 1 and 2 [124]
	Interleukin 8 [125]
	Matrix-degrading enzymes [110, 126]

for the increased levels found in the serum and urine of bladder cancer patients [97].

Angiogenesis inhibitors

While much of the research on bladder cancer angiogenesis to date has focused on inducers, we know, based on the balance hypothesis, that this represents only part of the puzzle of predicting a tumor's metastatic potential. Many inhibitors of angiogenesis exist, including thalidomide [98], interleukin-12 [99], angiostatin [100], and thrombospondin-1 [101, 102]. While angiostatin has been shown to inhibit the growth of Lewis lung carcinoma [100], human breast cancer [103], human colon cancer [103], and human prostate cancer [104] in animal models, only thrombospondin-1 has been examined in human bladder cancer.

Thrombospondin-1 (TSP) is an extracellular matrix glycoprotein that has been shown to be a potent inhibitor of angiogenesis, both *in vitro* and *in vivo* [105, 106]. Campbell and colleagues showed that conditioned medium from normal urothelial cells contained high levels of TSP-1 and could inhibit angiogenesis induced by VEGF and bFGF in bladder cancer cell lines.

Furthermore, they showed that the inhibitory activity of TSP-1 could be relieved by neutralizing antibody to TSP-1 [88]. Our group reported that TSP expression can be determined using antigen retrieval immunohistochemistry in routinely processed formalin-fixed, paraffin-embedded tissue [107]. Employing this technique, we evaluated 163 patients with invasive bladder cancer for TSP expression. Patients with low TSP expression exhibited higher recurrence rates and decreased overall survival when compared to patients with moderate or high TSP expression. This association was strongest in those patients with organ-confined disease. Furthermore, TSP expression remained an independent predictor of both disease recurrence and overall survival in the presence of tumor stage, histologic grade and lymph node status. In addition, in this same cohort of patients, we found that tumors with a low TSP expression were significantly more likely to demonstrate high microvessel density counts [108].

The extracellular matrix and angiogenesis

The ability of a tumor to invade surrounding stroma is one hallmark of metastasis. Stromal-epithelial interactions and matrix degrading enzymes undoubtedly play a role in the tumor's ability to invade. The composition of the ECM serves to maintain endothelial cell function, and provides a scaffolding through which the endothelium may attach and migrate during capillary formation. Joseph and colleagues have shown that bladder cancer cells can induce the production of scatter factor (a known angiogenesis inducer) by the underlying stromal cells [109]. Matrix metalloproteinases (MMP) are also

thought to play an important role in the degradation of the ECM. MMP-2 and MMP-9 are elevated in the serum and urine of patients with muscle-invasive bladder cancer, and are correlated with poorer disease free survival [110]. Furthermore, MMP-9 has an increased expression in TCC when compared to normal urothelium, and also correlated with increasing tumor stage [111].

Anti-angiogenic therapy

As a result of the improved understanding of tumor angiogenesis, clinical trials have now begun for some solid tumors in an attempt to develop effective anti-angiogenic therapies. Targeting the activated vessels associated with neovascularization of solid tumors provides several advantages over conventional forms of treatment. First, all blood vessels are readily accessible via the circulation, and are required by all tumors for growth and metastasis, yet are not necessary for normal physiologic function except for wound healing and female fertility [112]. Secondly, the endothelium targeted by an anti-angiogenic approach is non-neoplastic and maintains its full complement of regulatory mechanisms. Endothelial cells are unlikely to undergo the changes found in solid tumors that allow for the development of drug resistance [113]. Furthermore, there is a paucity of side effects associated with anti-angiogenic therapy—no bone marrow suppression or gastrointestinal mucosal alteration [114] making it a more desirable form of treatment than conventional chemotherapy.

Anti-VEGF therapy, consisting of humanized monoclonal antibodies directed at the VEGF protein, has demonstrated anti-tumor activity in animals [115]. All hereditary and most sporadic (clear cell) renal cell carcinomas (RCC) are associated with a defect in the VHL tumor suppressor gene located on chromosome 3p. A function of the VHL gene regulates the expression of proteins, including suppressing the expression of VEGF. Following a mutation in the gene or a partial deletion of chromosome 3, VEGF overexpression occurs and is thought to contribute to the many vascular manifestations characteristic of VHL syndrome [116]. Anti-VEGF therapy is now under investigation as a novel therapy against metastatic renal cell carcinoma [117].

While no clinical trial utilizing anti-angiogenic agents in bladder cancer has yet been established, the potential benefits are obvious. Anti-angiogenic therapy could be used for chemoprevention in patients at high risk for recurrence or progression, possibly through intravesical administration. A second indication would be for adjuvant therapy in patients at high risk for progression (advanced tumor stage, regional nodal involvement, altered p53) following radical cystectomy. This is particularly appealing, as the tumor burden is relatively low, and experimental models suggest that maintaining micrometastases in a dormant state for prolonged periods of time may be a reasonable goal, especially for older patients. Younger patients would likely require long-term or

intermittent administration of anti-angiogenic agents to maintain a disease-free or disease-stable state [118].

There is now ample evidence to suggest that bladder cancer is, in part, dependent on angiogenesis for growth and metastasis. However, angiogenesis remains a complex, tightly coordinated process not yet fully understood. While recent work has suggested that upregulation of inducers such as bFGF and VEGF, and/or downregulation of inhibitors like TSP-1 are important in determining a tumor's angiogenic phenotype, we are probably only looking at the tip of the iceberg in understanding how complex cellular interactions lead to tumor progression and metastasis. While therapies directed at individual inducers hold promise, this complexity of the angiogenic process makes it unlikely that targeting a single inducer will be an adequate treatment. Nevertheless, improved understanding of the molecular pathways regulating angiogenesis will undoubtedly improve tumor prognostication and treatment options.

Future directions

For now, it is clear that conventional histopathologic evaluation of bladder cancer, including determination of tumor grade and stage, is inadequate to accurately predict the behavior of many bladder cancers. One goal of understanding the biology of urothelial cell transformation and disease progression is to provide diagnostic markers that will help predict a tumor's natural history as well as its response to treatment. As we begin to decipher the molecular events that determine malignant transformation, cellular proliferation, and ultimately metastasis, we have come to realize the incredible complexity of bladder cancers. As a result, it is becoming obvious that diagnostic and therapeutic capabilities cannot be based on the knowledge of a single gene or protein, but rather on a complete understanding of chromosomal alterations, cell cycle activity, and extracellular interactions. Still, we are beginning to witness the emergence of molecular markers that provide some information on an individual tumor's biologic potential. Initially, these markers will help the transition from molecular science to clinical application, guiding therapeutic treatment plans for individual patients. This will indeed herald a new era, integrating molecular biology with surgery, pathology, and medical oncology for the purpose of disease management. Ultimately, this integration will require well designed prospective clinical trials of tumor markers and therapeutic drugs, if medicine is to truly benefit from this "molecular revolution". Thus, it behooves all urologists, clinicians and scientists alike who are involved in bladder cancer management to gain an understanding of urothelial cell tumor biology, so that there is an adequate knowledge of the importance of these trials. This will improve patient accrual, decrease the time necessary to obtain definitive results, and eventually allow for improved patient care and outcomes.

References

1. Cancer Statistics (2003)
2. Skinner DG, Lieskovsky G (1998) 16 years experience in the management of patients with deeply invasive bladder cancer. *Eur Urol* 14: 30
3. Droller MJ (1998) Markers in bladder cancer—issues to consider. *JUrol* 160: 2009
4. Malkowicz SB (1997) Superficial bladder cancer: the role of molecular markers in the treatment of high-risk superficial disease. *Semin Urol Oncol* 15: 169
5. Adishead JM, Kessler AM, Ogden CW (1998) Genetic initiation, progression and prognostic markers in transitional cell carcinoma of the bladder: a summary of the structural and transcriptional changes, and the role of developmental genes. *Br JUrol* 82: 503
6. Kroft SH, Oyasu R (1994) Urinary bladder cancer: mechanisms of development and progression. *Lab Invest* 71: 158
7. Moriyama N, Umeda T, Akaza H et al. (1989) Expression of ras p21 oncogene product on human bladder tumors. *Urol Int* 44: 260
8. Nagata Y, Abe M, Kobayashi K et al. (1990) Point mutations of c-ras genes in human bladder cancer and kidney cancer. *Jpn J Cancer Res* 81: 22
9. Fradet Y (1992) Markers of prognosis in superficial bladder cancer. *Semin Urol* 10: 28
10. Fontana D, Bellina M, Scoffone C et al. (1996) Evaluation of c-ras oncogene product (p21) in superficial bladder cancer. *Eur Urol* 29: 470
11. Koskinen PJ, Alitalo K (1993) Role of myc amplification and overexpression in cell growth, differentiation and death. *Semin Cancer Biol* 4: 3
12. Berns EM, Klijn JG, Van Putten WL et al. (1992) c-myc amplification is a better prognostic factor than HER2/neu amplification in primary breast cancer. *Cancer Res* 52: 1107
13. Kotake T, Saiki S, Kinouchi T et al. (1990) Detection of the c-myc gene product in urinary bladder cancer. *Jpn J Cancer Res* 81: 1198
14. Watt RA, Shatzman AR, Rosenberg M (1985) Expression and characterization of the human c-myc DNA-binding protein. *Mol Cell Biol* 5: 448
15. Lipponen PK (1995) Expression of c-myc protein is related to cell proliferation and expression of growth factor receptors in transitional cell bladder cancer. *J Pathol* 175: 203
16. Underwood M, Bartlett J, Reeves J et al. (1995) C-erbB-2 gene amplification: a molecular marker in recurrent bladder tumors? *Cancer Res* 55: 2422
17. Akiyama T, Sudo C, Ogawara H et al. (1986) The product of the human c-erbB-2 gene: a 185-kilodalton glycoprotein with tyrosine kinase activity. *Science* 232: 1644
18. Lee J, Dull TJ, Lax I. et al (1989) HER2 cytoplasmic domain generates normal mitogenic and transforming signals in a chimeric receptor. *EMBO J* 8: 167
19. Slamon DJ, Godolphin W, Jones LA et al. (1989) Studies of the HER-2/neu protooncogene in human breast and ovarian cancer. *Science* 244: 707
20. Gorgoulis VG, Barbatis C, Poulias I et al. (1995) Molecular and immunohistochemical evaluation of epidermal growth factor receptor and c-erbB-2 gene product in transitional cell carcinomas of the urinary bladder: a study in Greek patients. *Mod Pathol* 8: 758
21. Sato K, Moriyama M, Mori S et al. (1992) An immunohistologic evaluation of C-erbB-2 gene product in patients with urinary bladder carcinoma. *Cancer* 70: 2493
22. Moriyama M, Akiyama T, Yamamoto T et al (1991) Expression of c-erbB-2 gene product in urinary bladder cancer. *J Urol* 145: 423
23. Lipponen P, Eskelinen M (1994) Expression of epidermal growth factor receptor in bladder cancer as related to established prognostic factors, oncoprotein(c-erbB-2, p53) expression and long-term prognosis. *Br J Cancer* 69: 1120

24. Mellon JK, Lunec J, Wright C et al. (1996) C-erbB-2 in bladder cancer: molecular biology, correlation with epidermal growth factor receptors and prognostic value [see comments]. *J Urol* 155: 321
25. Li M, Zhang ZF, Reuter VE et al. (1996) Chromosome 3 allelic losses and microsatellite alterations in transitional cell carcinoma of the urinary bladder. *Am J Pathol* 149: 229
26. Takle LA, Knowles MA (1996) Deletion mapping implicates two tumor suppressor genes on chromosome 8p in the development of bladder cancer. *Oncogene* 12: 1083
27. Wagner U, Bubendorf L, Gasser TC et al. (1997) Chromosome 8p deletions are associated with invasive tumor growth in urinary bladder cancer. *Am J Pathol* 151: 753
28. Habuchi T, Devlin J, Elder PA et al. (1995) Detailed deletion mapping of chromosome 9q in bladder cancer: evidence for two tumour suppressor loci. *Oncogene* 11: 1671
29. Keen AJ, Knowles MA. (1994) Definition of two regions of deletion on chromosome 9 in carcinoma of the bladder. *Oncogene* 9: 2083
30. Simoneau AR, Spruck CH3rd, Gonzalez-Zulueta M et al. (1996) Evidence for two tumor suppressor loci associated with proximal chromosome 9p to q and distal chromosome 9q in bladder cancer and the initial screening for GAS 1 and PTC mutations. *Cancer Res* 56: 5039
31. Knowles MA (1999) The genetics of transitional cell carcinoma: progress and potential clinical application. *BJU Int* 84: 412
32. Orlov I, Lacombe L, Hannon GJ et al. (1995) Deletion of the p16 and p15 genes in human bladder tumors [see comments]. *J Nat Cancer Inst* 87: 1524
33. Packenham JP, Taylor JA, Anna CH et al. (1995) Homozygous deletions but no sequence mutations in coding regions of p 15 or p16 in human primary bladder tumors. *Mol Carcinogen* 14: 147
34. Williamson MP, Elder PA, Shaw ME et al. (1995) p16(CDKN2) is a major deletion target at 9p21 in bladder cancer. *Human Mol Genet* 4: 1569
35. Wu Q, Possati L, Montesi M et al. (1996) Growth arrest and suppression of tumorigenicity of bladder-carcinoma cell lines induced by the P16/CDKN2(p16INK4A, MTS1) gene and other loci on human chromosome 9. *Int J Cancer* 65: 840
36. Habuchi T, Luscombe M, Elder PA et al. (1988) Structure and methylation-based silencing of a gene(DBCCR1) within a candidate bladder cancer tumor suppressor region at 9p32-q33. *Genomics* 48: 277
37. Habuchi T, Ogawa O, Kakehi Y et al. (1993) Accumulated allelic losses in the development of invasive urothelial cancer. *Int J Cancer* 53: 579
38. Olumi AF, Tsai YC, Nichols PW et al. (1990) Allelic loss of chromosome 17p distinguishes high grade from low grade transitional cell carcinomas of the bladder. *Cancer Res* 50: 7081
39. Sidransky D, Von Eschenbach A, Tsai YC et al. (1991) Identification of p53 gene mutations in bladder cancers and urine samples. *Science* 252: 706
40. Williamson MP, Elder PA, Knowles MA (1994) The spectrum of TP53 mutations in bladder carcinoma. *Genes, Chromosomes Cancer* 9: 108
41. Cordon-Cardo C, Dalbagni G, Saez GT et al. (1994) p53 mutations in human bladder cancer: genotypic versus phenotypic patterns. *Int J Cancer* 56: 347
42. Esrig D, Spruck CH, Nichols PW et al. (1993) p53 nuclear protein accumulation correlates with mutations in the p53 gene, tumor grade, and stage in bladder cancer. *Am J Pathol* 143: 1389
43. Mareel MM, Van Roy FM, Bracke ME (1993) How and when do tumor cells metastasize? *Crit Rev Oncogen* 4: 559
44. Sparkes RS, Sparkes MC, Wilson MG et al. (1980) Regional assignment of genes for human esterase D and retinoblastoma to chromosome band 13814. *Science* 208: 1042
45. Logothetis CJ, Xu HJ, Ro JY et al. (1992) Altered expression of retinoblastoma protein and known prognostic variables in locally advanced bladder cancer [see comments]. *J Nat Cancer Inst* 84: 1256
46. Cordon-Cardo C, Wartinger D, Petrylak D et al. (1992) Altered expression of the retinoblastoma gene product: prognostic indicator in bladder cancer [see comments]. *J Nat Cancer Inst* 84: 1251
47. Xu HJ, Cairns P, Hu SX et al. (1993) Loss of RB protein expression in primary bladder cancer correlates with loss of heterozygosity at the RB locus and tumor progression. *Int J Cancer* 53: 781
48. Markl ID, Salem CE, Jones PA (2000) Molecular biology of bladder cancer. In: Vogelzang N, Scardino PT, Shipley WU, Coffey DS (eds) *Comprehensive textbook of genitourinary oncology* (). Lippincott Williams and Wilkins, Philadelphia, p 298
49. Markl ID, Jones PA (1998) Presence and location of TP53 mutation determines pattern of CDKN2A/ARF pathway inactivation in bladder cancer. *Cancer Res* 58: 5348
50. Spruck CH3rd, Ohneseit PF, Gonzalez-Zulueta M, et al. (1994) Two molecular pathways to transitional cell carcinoma of the bladder. *Cancer Res* 54: 784
51. Cordon-Cardo C (1995) Mutations of cell cycle regulators. Biological and clinical implications for human neoplasia. *Am J Pathol* 147: 545
52. Fung YK, Murphree AL, T'Ang A et al. (1987) Structural evidence for the authenticity of the human retinoblastoma gene. *Science* 236: 1657
53. Presti JCJr., Reuter VE, Galan T et al. (1991) Molecular genetic alterations in superficial and locally advanced human bladder cancer. *Cancer Res* 51: 5405
54. Cote RJ, Chatterjee SJ (1999) Molecular determinants of outcome in bladder cancer. *Cancer J Sci Am* 5: 2
55. Ishikawa J, Xu HJ, Hu SX et al. (1991) Inactivation of the retinoblastoma gene in human bladder and renal cell carcinomas. *Cancer Res* 51: 5736
56. Hollstein M, Sidransky D, Vogelstein B et al. (1991) p53 mutations in human cancers. *Science* 253: 49
57. Lane DP (1992) Cancer. p53, guardian of the genome [see comments]. *Nature* 358: 15
58. Vet JA, Bringuier PP, Schaafsma HE et al. (1995) Comparison of P53 protein overexpression with P53 mutation in bladder cancer: clinical and biologic aspects. *Lab Invest* 73: 837
59. Dalbagni G, Cordon-Cardo C, Reuter V et al. (1995) Tumor suppressor gene alterations in bladder carcinoma. Translational correlates to clinical practice. *Surg Oncol Clinics North Am* 4: 231
60. Esrig D, Elmajian D, Groshen S et al. (1994) Accumulation of nuclear p53 and tumor progression in bladder cancer [see comments]. *N Engl J Med* 331: 1259
61. Sarkis AS, Dalbagni G, Cordon-Cardo C et al (1993) Nuclear overexpression of p53 protein in transitional cell bladder carcinoma: a marker for disease progression. *J Nat Cancer Inst* 85: 53
62. Greenblatt MS, Bennett WP, Hollstein M et al. (1994) Mutations in the p53 tumor suppressor gene: clues to cancer etiology and molecular pathogenesis. *Cancer Res* 54: 4855
63. Cote RJ, Dunn MD, Chatterjee SJ et al. (1998) Elevated and absent pRb expression is associated with bladder cancer progression and has cooperative effects with p53. *Cancer Res* 58: 1090
64. Cordon-Cardo C, Zhang ZF, Dalbagni G et al. (1997) Cooperative effects of p53 and pRb alterations in primary superficial bladder tumors. *Cancer Res* 57: 1217
65. Parker SB, Eichele G, Zhang P et al. (1995) p53-independent expression of p21Cip1 in muscle and other terminally differentiating cells [see comments]. *Science* 267: 1024
66. Stein JP, Ginsberg DA, Grossfeld GD et al. (1998) Effect of p21WAF1/CIP1 expression on tumor progression in bladder cancer [see comments]. *J Nat Cancer Inst* 90: 1072
67. Serrano M, Hannon GJ, Beach D (1993) A new regulatory motif in cell-cycle control causing specific inhibition of cyclin D/CDK4 [see comments]. *Nature* 366: 704

68. Gonzalez-Zulueta M, Bender CM, Yang AS et al. (1995) Methylation of the 5' CpG island of the p16/CDKN2 tumor suppressor gene in normal and transformed human tissues correlates with gene silencing. *Cancer Res* 55: 4531
69. Chin L, Pomerantz J, DePinho RA (1998) The INK4a/ARF tumor suppressor: one gene-two products-two pathways. *Trends Biochem Sci* 23: 291
70. Quelle DE, Zindy F, Ashmun RA et al. (1995) Alternative reading frames of the INK4a tumor suppressor gene encode two unrelated proteins capable of inducing cell cycle arrest. *Cell* 83: 993
71. Orlow I, LaRue H, Osman I et al. (1999) Deletions of the INK4A gene in superficial bladder tumors. Association with recurrence. *Am J Pathol* 155: 105
72. Pomerantz J, Schreiber-Agus N, Liegeois NJ et al. (1998) The Ink4a tumor suppressor gene product, p19Arf, interacts with MDM2 and neutralizes MDM2's inhibition of p53. *Cell* 92: 713
73. Momand J, Wu HH, Dasgupta G (2000) MDM2—master regulator of the p53 tumor suppressor protein. *Gene* 242: 15
74. Lianes P, Orlow I, Zhang ZF et al. (1994) Altered patterns of MDM2 and TP53 expression in human bladder cancer [see comments]. *J Nat Cancer Inst* 86: 1325
75. Folkman J (1985) Tumor angiogenesis. *Adv Cancer Res* 43: 175
76. Folkman J (1992) The role of angiogenesis in tumor growth. *Semin Cancer Biol* 3: 65
77. Hanahan D, Folkman J (1996) Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. *Cell* 86: 353
78. Volpert OV, Dameron KM, Bouck N (1997) Sequential development of an angiogenic phenotype by human fibroblasts progressing to tumorigenicity. *Oncogene* 14: 1495
79. Polverini PJ (1995) The pathophysiology of angiogenesis. *Crit Rev Oral Biol Med* 6: 230
80. Blood CH, Zetter BR (1990) Tumor interactions with the vasculature: angiogenesis and tumor metastasis. *Biochim Biophys Acta* 1032: 89
81. Folkman J (1987) What is the role of angiogenesis in metastasis from cutaneous melanoma? *Eur J Cancer Clin Oncol* 23: 361
82. Weidner N, Folkman J, Pozza F et al. (1992) Tumor angiogenesis: a new significant and independent prognostic indicator in early-stage breast carcinoma [see comments]. *J Nat Cancer Inst* 84: 1875
83. Weidner N, Carroll PR, Flax J et al. (1993) Tumor angiogenesis correlates with metastasis in invasive prostate carcinoma. *Am J Pathol* 143: 401
84. Barnhill RL, Piepkorn MW, Cochran AJ et al. (1998) Tumor vascularity, proliferation, and apoptosis in human melanoma micrometastases and macrometastases [see comments]. *Arch Dermatol* 134: 991
85. Bochner BH, Cote RJ, Weidner N et al. (1995) Angiogenesis in bladder cancer: relationship between microvessel density and tumor prognosis. *J Nat Cancer Inst* 87: 1603
86. Jaeger TM, Weidner N, Chew K et al. (1995) Tumor angiogenesis correlates with lymph node metastases in invasive bladder cancer. *J Urol* 154: 69
87. Dickinson AJ, Fox SB, Persad RA et al. (1994) Quantification of angiogenesis as an independent predictor of prognosis in invasive bladder carcinomas. *Br J Urol* 74: 762
88. Campbell SC, Volpert OV, Ivanovich M et al (1998) Molecular mediators of angiogenesis in bladder cancer. *Cancer Res* 58: 1298
89. Chodak GW, Scheiner CJ, Zetter BR (1981) Urine from patients with transitional-cell carcinoma stimulates migration of capillary endothelial cells. *N Engl J Med* 305: 869
90. Nguyen M, Watanabe H, Budson AE et al (1993) Elevated levels of the angiogenic peptide basic fibroblast growth factor in urine of bladder cancer patients. *J Nat Cancer Inst* 85: 241
91. O'Brien TS, Harris AL (1995) Angiogenesis in urological malignancy. *Br J Urol* 76: 675
92. Bochner B, McHugh R, Spitz A, Groshen S, Skinner DG, Nichols PW (1997) Basic fibroblast growth factor (bFGF) in bladder cancer: elevated urinary levels predict pathologic stage. *J Urol* 157: 341
93. Crew JP, O'Brien T, Bicknell R et al (1999) Urinary vascular endothelial growth factor and its correlation with bladder cancer recurrence rates [see comments]. *J Urol* 161: 799
94. Williams SG, Feng A, Skinner DG, Nichols PW, Bochner BH (2000) Urine levels of vascular endothelial growth factor predict recurrence in bladder cancer. *J Urol* 163: 133
95. O'Brien T, Cranston D, Fuggle S et al. (1995) Different angiogenic pathways characterize superficial and invasive bladder cancer. *Cancer Res* 55: 510
96. Sato K, Sasaki R, Ogura Y et al. (1998) Expression of vascular endothelial growth factor gene and its receptor (flt-1) gene in urinary bladder cancer. *Tohoku J Exp Med* 185: 173
97. O'Brien T, Cranston D, Fuggle S et al. (1997) Two mechanisms of basic fibroblast growth factor-induced angiogenesis in bladder cancer. *Cancer Res* 57: 136
98. D'Amato RJ, Loughnan MS, Flynn E et al. (1994) Thalidomide is an inhibitor of angiogenesis. *Proc Nat Acad Sci U.S.A* 91: 4082
99. Voest EE, Kenyon BM, O'Reilly MS et al. (1995) Inhibition of angiogenesis in vivo by interleukin 12 [see comments]. *J Nat Cancer Inst* 87: 581
100. O'Reilly MS, Holmgren L, Shing Y et al. (1994) Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma [see comments]. *Cell* 79: 315
101. Sheibani N, Frazier WA (1998) Down-regulation of platelet endothelial cell adhesion molecule-1 results in thrombospondin-1 expression and concerted regulation of endothelial cell phenotype. *Mol Biol Cell* 9: 701
102. Bochner B, Windham CQ, Pao MM, Kasahara N, Skinner DG, Jones PA (1998) Overexpression of thrombospondin-1 inhibits bladder tumor growth: a novel therapy for invasive bladder cancer. *J Urol* 159: 282
103. Saito T, Kimura M, Kawasaki T et al. (1996) Correlation between integrin alpha 5 expression and the malignant phenotype of transitional cell carcinoma. *Br J Cancer* 73:
104. O'Reilly MS, Holmgren L, Chen C et al. (1996) Angiostatin induces and sustains dormancy of human primary tumors in mice. *Nat Med* 2: 689
105. Iruela-Arispe ML, Bornstein P, Sage H (1991) Thrombospondin exerts an antiangiogenic effect on cord formation by endothelial cells in vitro. *Proc Nat Acad Sci U.S.A* 88: 5026
106. Good DJ, Polverini PJ, Rastinejad F et al. (1990) A tumor suppressor-dependent inhibitor of angiogenesis is immunologically and functionally indistinguishable from a fragment of thrombospondin. *Proc Nat Acad Sci U.S.A* 87: 6624
107. Grossfeld GD, Shi SR, Ginsberg DA et al. (1996) Immunohistochemical detection of thrombospondin-1 in formalin-fixed, paraffin-embedded tissue. *J Histochem Cytochem* 44: 761
108. Grossfeld GD, Ginsberg DA, Stein JP et al (1997) Thrombospondin-1 expression in bladder cancer: association with p53 alterations, tumor angiogenesis, and tumor progression. *J Nat Cancer Inst* 89: 219
109. Joseph A, Weiss GH, Jin L et al. (1995) Expression of scatter factor in human bladder carcinoma. *J Nat Cancer Inst* 87: 372
110. Gohji K, Fujimoto N, Fujii A et al. (1996) Prognostic significance of circulating matrix metalloproteinase-2 to tissue inhibitor of metalloproteinases-2 ratio in recurrence of urothelial cancer after complete resection. *Cancer Res* 56: 3196
111. Gohji K, Fujimoto N, Ohkawa J et al. (1998) Imbalance between serum matrix metalloproteinase-2 and its inhibitor as a predictor of recurrence of urothelial cancer. *Br J Cancer* 77: 650
112. Folkman J (1995) *Seminars in Medicine of the Beth Israel Hospital, Boston. Clinical applications of research on angiogenesis* [see comments]. *N Engl J Med* 333: 1757

113. Boehm T, Folkman J, Browder T et al. (1997) Antiangiogenic therapy of experimental cancer does not induce acquired drug resistance [see comments]. *Nature* 390: 404
114. Folkman J (1997) Angiogenesis and angiogenesis inhibition: an overview. *Exs* 79: 1
115. Schlaeppli JM, Wood JM (1999) Targeting vascular endothelial growth factor(VEGF) for anti-tumor therapy, by anti-VEGF neutralizing monoclonal antibodies or by VEGF receptor tyrosine-kinase inhibitors. *Cancer Metastasis Rev* 18: 473
116. Siemeister G, Martiny-Baron G, Marme D (1998) The pivotal role of VEGF in tumor angiogenesis: molecular facts and therapeutic opportunities. *Cancer Metastasis Rev* 17: 241
117. Bichler KH, Wechsel HW (1999) The problematic nature of metastasized renal cell carcinoma. *Anticancer Res* 19: 1463
118. Campbell SC, Bochner BH (1998) Angiogenesis in bladder cancer. *Mol Urol* 2: 279
119. Chopin DK, Caruelle JP, Colombel M et al. (1993) Increased immunodetection of acidic fibroblast growth factor in bladder cancer, detectable in urine. *J Urol* 150: 1126
120. Ravary V, Jouanneau J, Gil Diez S, et al. (1992) Immunohistochemical detection of acidic fibroblast growth factor in bladder transitional cell carcinoma. *Urol Res* 20: 211
121. Crew JP, O'Brien T, Bradburn M et al. (1997) Vascular endothelial growth factor is a predictor of relapse and stage progression in superficial bladder cancer. *Cancer Res* 57: 5281
122. O'Brien TS, Fox SB, Dickinson AJ et al. (1996) Expression of the angiogenic factor thymidine phosphorylase/platelet-derived endothelial cell growth factor in primary bladder cancers. *Cancer Res* 56: 4799
123. Rosen EM, Goldberg ID (1997) Regulation of angiogenesis by scatter factor. *Exs* 79: 193
124. Eder IE, Stenzl A, Hobisch A et al. (1996) Transforming growth factors-beta 1 and Beta 2 in serum and urine from patients with bladder carcinoma [see comments]. *J Urol* 156: 953
125. Andrawis R, Contrino J, Lindquist RR et al. (1997) Interleukin-8 expression and human bladder cancer:in-situ and in-vitro expression of IL-8 by human bladder cancer cells. *J Urol* 157: 28
126. Gohji K, Fujimoto N, Komiyama T et al. (1996) Elevation of serum levels of matrix metalloproteinase-2 and -3 as new predictors of recurrence in patients with urothelial carcinoma. *Cancer* 78: 2379